

## Sample Preparation for DNA Methylation Analysis

### RRBS

(Use one of the following recommendations for sample submission.)

#### I. Prepare frozen pellets from cell cultures

1. Grow  $1 \times 10^5$  to  $5 \times 10^6$  cells in culture.
2. Transfer cell culture to a conical tube. (If cells are adherent, scrape them thoroughly from the culture surface prior to transferring to a conical tube).
3. Centrifuge tubes at  $800 \times g$  in a refrigerated centrifuge for 5 minutes to pellet the cells. Decant culture media.
4. Resuspend cells in 10 ml chilled PBS by pipetting up and down, then spin again at  $800 \times g$  in a refrigerated centrifuge for 5 minutes to pellet the cells.
5. Decant PBS, freeze cell pellets on dry ice and store at  $-80^\circ\text{C}$ .

#### II. Freeze animal tissue

1. Remove an appropriate amount of tissue from the animal (20-200 mg for most tissues).
2. Place tissue in a 1.5 ml microfuge tube or 15 ml conical, freeze on dry ice and store at  $-80^\circ\text{C}$ .

#### III. Prepare DNA

1. Prepare genomic DNA from cell culture or animal tissue using a Qiagen QIAmp DNA Mini Kit (cat # 51304) or comparable genomic DNA isolation method.
2. Elute or resuspend DNA in 10 mM Tris, pH 8.
3. Run 100-200 ng of DNA on a 1% agarose gel to show high molecular weight DNA.
4. Send 100 ng to 10  $\mu\text{g}$  of DNA at a minimum concentration of 10 ng/ $\mu\text{l}$ .  
NOTE: Minimum concentration is required in order not to dilute reaction volume.